vet. Med. J. Giza 38, No. 1, 95-103 (1990).

IMMUNE RESPONSE OF CHICKEN INFECTED WITH QUARANFIL VIRUS

BY

A.G. HEGAZI AND KOUKA S, ABD EL WAHAB
National Research Centre and Faculty of Med. for
Girls, El Azhar Univ.

(Received: 2.12.1989)

INTRODUCTION

Quranfile virus (QRFV) is one of the unclassified group viruses of Arina viruses (Taylor et al., 1966) at the last few years it was believed that this virus is tick borne Arboviruses. It seems quite clear that birds constitute the basic vertebrate hsot, and that the maintenance cycle is from tick to birds and back to tick (Hoogstraal, 1973).

Studies employed in vitro correlates of cellular immunity have revealed further evidence of suppressed immne reactivity durring acute viral infection. The reactivity of lymphoid cells to antigen or phytohaemagglutinin (PHA) was decreased in animals naturally infected with viruses or vaccine induced as measles (Zweiman, 1971 and Zweiman et al., 1971), influenza (Reed et al., 1972) and hepatitis virus (Willems et al., 1969). On the other hand, Gimboro disease suppressed B cell mainly present in Bursa of Fabricous.

The aim of this investigation is to through a light on the role of QRFV on the chicken immune response.

Immune Response of Chicken infected with Quaranfil...

MATERIAL: AND METHODS

Sixty chicks (Dokki-4) about 14 days old were used Sixty chicks were obtained in this investigation. These chicks were obtained through the Anshas Research Station, Ministry of Agriculture. The chicks were divided into 6 gorups, Agriculture.

each of 10 The first group was injected intraperoeach of 10 line with 1 ml sheep red blood cells in a dose of 108/ml (Hegazi, 1979), the second group was injected I/P with bovine serum albumin (BAS) (Sigma Lab. U.S.A.) in a dose of 50 mg/ml (Hegazi et al., 1985 b), the third group was injected intramascular (I/M) with quaranfil virus (QRFV)(AR-1117 in suckling mous brain, Rocky Mountain Lab. U.S.A.) in a dose of 107.1 ISMBILD50/ml (Intracerabral suckling mous brain infective lethal dose fifty) (Kouka, 1978). The fourth group was injected I/P with SRBC (sheep red blood oells) 1 ml and I/M with 107.1 ISMBILD50/ml ORFV. The fifth group was injected I/P with BSA (50 mg.ml) & I/M with $10^{7.1}$ QRFV. While the sixth group was kept as normal control. All groups were kept under observation.

Five chicks from each group were taken after four days post treatment to collect heparnized blood and spleen. The whole blood culture and splenocytes were used to study the dose response curve (Hall and Gorden, 1976 & Hegazi, 1981). The plaque assay was embloyed on the splenocytes (Jerne and Nordin, 1963 and Hegazi, 1979) by using SRBC, SRBC conjugated with BSA and SRBC conjugated with QRFV as indicator cells for detection of PFC/106 splenocytes.

All groups after 14 days were injected intradermally (I/D) on the right foot pad (Hegazi et al., 1985 a) with its specific antigen, while the left once injected with normal saline and considered as control. The sixth group (control group) was injected in both measured (Goto et al., 1978) and recorded for detection of the foot pad index.

A.G. Hegazi and Kouka S. Abd El-Wahab.

RESULTS

Table, 1 shows the stimulation index (SI) (dose response curve) of whole blood culture and spleen lymphocyte cultures to PHA in the six groups.

- a. Whole blood culture: The normal stimulation indecies were 8.3, 6.3 and 5.1 at PHA dilutions 1:25, 1.75 and 1:225 respectivelly. It is noticed that a great reduction in the stimualtion index in chicks injected with SRBC, QRFV + SRBC, while an increase in chicks injected with BSA, ORFV + BSA and QRFV. The optimum PHA dilution was 1:225 which gaves the highest SI in groups infected with QRFV.
- b. Spleen lymphocyte culture: The SI of the normal spleen lymphocyte culture was 0.5, 6.9 and 1.3 at 1:25, 1:75 and 1:225 PHA dilutions respectively. There was slight increase in SI of spleen lymphocyte culture of chicks injected with SRBC, BSA, QRFV + SRBC and QRFV + BSA. While a great increase of SI in chicks infected with QRFV. The optimum PHA dilution was 1:225 specially in groups infected with QRFV.
- c. Foot pad index: Table, 2 demonstrated the delayed hypersensitivity (intradermal skin test) as a sequential reactivity post injection with each specific antigen as sensitized antigens. From this table it is clear that the highest foot pad index in chicks injected with BSA (2.0 mm), QRFV + BSA (1:1) and QRFV (1.0), while the lowest foot pad index in chicks injected I/D by SRBC (0.3 mm) and there are a moderate reaction in chicks injected with QRFV + SRBC (0.5 mm).
- d. Plaque forming cells: The PFC/10⁶ spleen lymphocytes was shown in the Table (2), it is clear that the highest PFC level was 440 in SRBC group, while a reduction in BSA, SRBC + QRFV, BSA + QRFV and a suppressive effect was observed in chicks infected with QRFV only (7).

Table (1): Stimulation index of chinken whole blood culture and spleen

		In			
Groups	FHA dilution	Whole bi	DPMCO	Spleen SI	cell culture DPM
	25	8.3	1336	0.5	254
	1:25	6.3	1021	6.9	15919
Control	1:75	5.1	75B	1.3	886
	1: 225	4.0	687	1.02	142
	1:25	2.3	. 292	3.4	6040
SRBC	1: 225	4.5	785	1.07	184
BSA	1:25	4.0	587	2.2	845
	1:75	6.0	919	1.6	400
	1: 225	12.0	2069	20.5	6962
	1:25	6.5	634	8.0	324
QRFV	1:75	4.5	415	7.0	7250
QIL T	1: 225	7.2	219	5.0	. 303
81W0 +	1 : 25	1.4	151	2.2	936
QRFV	1:75	1.5	169	7.1	4138
	1:225	2.8	679	2.2	1548
BSA +	1:25	3.4	1733	4.0	810
QRFY	1:75	3.9	1810	1.9	249
	1 : 225	6.2	736	.5.4	210
SI 1	Stimulation :	Index DPM	: Disentig	r ation]	per min.te
Table (2) Foot pad :	index and PPC	/ 10 ⁶ splee:	lymphod	oytes in chick

Test Control	SRac	BSA	QPFV	SRBC+QRFV	BSA+QRF
Foot pad index 0.0	0.3	2.0	1.0	0.5	1;1
PFC/10 ⁶ spleen 35 cells 96 hours	440	100	7	200	54

Immune Response of Chicken infected with Quaranfil...

DISCUSSION

Regarding the results of the stimulation index (mitogenic dose respons curve) of whole blood culture and spleen lymphocyte cultures revealed that the PHA used in the experiment induced a noticeable blastformation in all PHA dilutions. These results proved that the cells (whole blood & spleen lymphocyte cultures)under examination are functioning which concided with observations obtained by Kirohner and Blaese (1973) and Hegazi (1981) who reported that peripheral blood lymphocytes, splenic lymphocytes, bone marrow cells and even thymus cells exposed to PHA will developed blastformation.

The optimum stimulation of lymphocytes to PHA varied depending on the dose of PHA, yet it was the first aim to determine such parameter. It was noticed that optimum PHA dilution was 1:225 in all groups under the examination. Also many authers reached different optimum stimulation using different PHA dilution as Lee (1975); Lee (1978) and Hegazi (1981) who found a significant increase in PHA response up to 270 fold using 20 ug PHA, while Maheswaran and Thies (1975) used Con A at dilution of 0.4 ug and pokweed 80 ug at a cell concentration 2 x 10^6 cells.

Concerning the delayed hypersensitivity skin test as a sequential reactivity post injection with each (sensitized antigen) specific antigen revealed that the highest foot pad index in chicks injected with BSA, QRFV + BSA, while the lowest foot pad index was observed in chicks injected with SRBC and a moderate observed in QRFV. The rise of foot pad index indicareaction in QRFV. The rise of foot pad due to ted that there was an increase in foot pad due to stimulation with specific antigen. These findings stimulation with specific antigen. These findings also observed by many authers used different specific antigens in chickens as purified protein derivative antigens in chickens as purified protein derivative (PPD) of tuberculine (Cheville and Richards, 1971);

A.G. Hegazi and Kouka S. Abd El-Wahab.

Newcastle disease virus (Cheville and Beard, 1972 & Hegazi, 1981); PHA (Goto et al., 1978 and Mac Corkle et al., 1979); BSA (Hegazi et al., 1985 a) and Reo virus (Hegazi et al., 1989).

In order to apply the Jerne test for the enumeration of the antibody producing cells after inoculation of the antibody project (QRFV & BSA), it was importwith dillerent antigen of virus and BSA that can be ant to prepare to carry the sheep red blood cells which used as an indicator. Conjugation of such antigen to red cells was the bases of Jerne and Nordin, (1963) for enumeration of PFC. The detection of the PFC at the 4th day post injection of different antigens was based of the data obtained by many authors as Eveland (1964); Abramoff and Brien (1968); Martin and Leslie (1974); Romaninkawa (1974); Hegazi (1979) and Hegazi et al., (1985 b) who found that the PFC appeared after the inoculation of antigen. The number of the PFC was usually greater at the fourth day post inoculation. It was clear that the difference in numbers of PFC/106 spleen cells in all groups under investigation may be due to the difference in the antigens nature.

From overmentianed data it can be concluded that the QRFV showed an immunosuppressive effect on chicken immune response detected by delayed hypersensitivity and PFC as well as deminished the non specific stimulation with PHA. Also the optimum dilution of PHA used in the stimulation of whole blood and spleen cell cultures was 1:225. in QRFV infected chicks.

SUMMARY

The immune response of chicken infected with Quaranfil (QRFV) virus was investigated. The dose response curve of whole blood culture and spleen lymphocyte cultures were deteated by non specific stimulation with phytomaemagglutinin (PHA) at different dilutions. The optimium stimulation was (1:225) with 0.5 x 10^6 cells in

Immune Response of Chicken infected with Quaranfil ...

whole blood culture, also (1:225) in spleen lymphocyte culture. The delayed hypersensitivity skin test as a sequential reactivity post injection with specific antigen revealed moderate reaction against QRFV. The plaque forming cells (PFC) was reduced in case of QRFV if it is compared with the control group (sheep red blood cells).

REFERENCES

- Abramoff, P. and Brian, N.B. (1986): Immunol. 100: 1204-1214.
- Cheville, N.P. and Beard, C.W. (1972): Lab. Invest.
 129-143.
- 3. Cheville, N.P. and Richards, S.W.D. (1971): Am.J. Pathol. 64: 97-117.
- 4. Eveland, W.C. (1964): J. Bact. 38: 1476-1481.
- 5. Goto, N.; Kodama, H.; Okado, K. and Fujinoto, Y. (1978): Poultry Sci. 57: 246-250.
- Hall, L.S. and Gordon, D.S. (1976): Immunol. Moth. 12: 31-38.
- 7. Hegazi, A.G. (1979): M.V.Sc.; Faculty of Vet. Med. Cairo Univ.
- 8. Hegazi, A.G. (1981): Ph.D. V.Sc.; Faculty of vet. Med. Cairo Univ.
- Hegazi, A.G.; Soufy, H.; Edriss, B.M. and Abd El-Haim, (1987): First Int. Conf. App. Sci. Vol. 3: 299-309.

- A.G. Hegazi and Kouka S. Abd El-Wahab.
- 10. Hegazi, A.G.; Shawki, M.M.; Arbid, M.S.; Soufy, H. and ElShabrawy, O. (1985 b): Egypt. J. Vet. Sci. 22 (2): 191-200.
- 11. Hegazi, A.G.; El Sanousi, A. and Kheir El Din, A.W. (1989): J. Egypt. Vet. Med. Ass. 49 (1 & 2): 731-740.
- 12. Hoogstraal, H. (1973): Virus and ticks, In: Viruses and invertebrate, A.J. Gibbs (ed) North Holland Publishing Co. P. 394.
- 13. Jerne, N.K. and Nordin, A.A. (1963): Sci. 140: 405-407.
- Kirohner, H. and Blaese, R.M. (1973): Exptl. Med. 138: 812-834.
- Kouka, S. Abd El-Wahab (1978): J. Egypt. Vet. Med. Ass. 38 (1) 83-90.
- 16. Lee, F. Loucy (1975): Avian Dis .18: 602-609.
- 17. Lee, F. Loucy (1978): Avian Dis. 22: 296-307.
- Maheswaran, S.K. and Thies, E.S., (1975): Am. J. Vet. Res. 63: 1053-1055.
- Martin, L.N. and Leslie, G.A. (1974): Immunol. 113: 123-126.
- Mc Corkle, F.M.; Stinson, Jr. R. and Glick, P. (1979): Cell Immunol. 46: 208-212.
- Reed, W.P.; Olds, J.W. and Kisch, A.L. (1972):
 J. Infect. Dis. 125: 398-400.
- 22. Romaninkawa, K.R. (1974): J. Bull. Vet. Inst. Pulawy, 18: 104-108.

Immune Response of Chicken infected with Quaranfil...

- 23. Taylor, R.M.; Hurlbut, H.S.; Work, T.H.; Kongston, T.P. and Hoogstraal, H. (1966): Am. J. Trop. Med. and Hyg. 15: 76-82.
- 24. Willems, F.T.C.; Melnick, J.L.; and Rawis, W.E. (1969): J. Virol. 3: 451-456.
- 25. Zweiman, B. (1971): J. Immunol. 106: 1154-1156.
- 26. Zweiman, B.; Pappaglania, D. Maibach, C. and Hildreth, E.A. (1971): Int. Arch. Allergy 40: 834-838.