

## ESCHERICHIA COLI IN MEAT PRODUCTS

BY

A.M. DARWISH\*, Z.M. NIAZI\*\*, AND  
E.M. ZAKI\*\*

\* Faculty of Vet. Med. Cairo University.

\*\* Animal Health Research Institute, Dokki, Cairo.

(Received: 25.5.1991).

### INTRODUCTION

Escherichia coli in food has been associated with outbreaks of gastrointestinal disease since 1903 (Delepine, 1903).

The presence of different members of Enterobacteriaceae particularly Escherichia coli in meat products was attributed to the contamination of such products (El-Mossalami, 1958 and Kleeberger et al., 1980).

External contamination of raw meat is a constant possibility from the moment of bleeding until consumption. There are several potential sources of contamination by microorganisms. These include contact with hide, skin, feet, the gastrointestinal tract content, aqueous sources and the instruments used for dressing (Knives, saws, cleaners or hooks) and even air borne contamination in the processing and storage areas (Niskanen and Pohja 1977 Agres et al., 1980, Stolle, 1981 and Jawetz et al., 1982).

In addition, meat products may be contaminated with Escherichia coli from food handlers, food utensils, soils and water under incomplete hygienic circumstances during manufacturing, packing and marketing of these products (Frazier and Westhoff, 1978).

## *Escherichia coli* in meat products.

The native habitat for *Escherichia coli* is the intestinal tract of man and animals; therefore its presence in foods generally indicates direct or indirect pollution of faecal origin. *Escherichia coli* is the classical indicator of the possible presence of enteric pathogens in foods.

On the other hand, meat products constitute a public health hazard either due to the presence of spoilage bacteria responsible for unfavourable changes, or pathogenic bacteria like *Escherichia coli* leading to harmful effects as infection or intoxication in human consumers. i.e. food borne infections (Mastievskii et al., 1971; Anon, 1978; Mehlman and Bomero, 1982).

Enteropathogenic *Escherichia coli* (EPEC) of certain serovars are well recognized pathogens as caused of infantile diarrhoea and/or gastrointestinal illness in adult humans (Broy, 1945; Dupont et al., 1971; Edelman and Levine, 1983).

*Escherichia coli* may also cause peritonitis, meningitis, enteritis, cystitis, pyelitis, pylonephritis, angiocholitis, salpingoophoritis, appendicitis, optotitis and purepeareal sepsis (Pyathin and Krivoshein 1980).

The present work was planned to study the following:

1. Incidence of *E. coli* in meat products using two techniques; direct plate count technique and multiple tube fermentation technique.
2. Biochemical and serological identification of *E. coli*.

*A.M. Darwish et al.*

## MATERIAL AND METHODS

100 samples, 25 each from fresh minced meat, fresh sausage, frozen beef burger and basterma slices were collected from Cairo and Giza markets, transported in cool cabinet and examined bacteriologically.

The samples were prepared according to the technique recommended by ICMSF (1978).

1. Determination of *Escherichia coli* count by the surface spread plate method recommended by Barraud et al. (1967) as well as by the multiple tube fermentation technique recommended by MLG (1974).
2. Isolation, biochemical and seriological identification of enteropathogenic strains of *E. coli* according to Cruickshank et al., 1975.

## RESULTS AND DISCUSSION

From Table (1), it is evident that *E. coli* isolates were detected in 26 (26%) of the examined samples. The incidence of *E. coli* in minced meat, sausage, beef burger and basterma was 44%, 40%, 12% and 8% respectively.

Using the direct plate count technique (DPC) a total of 17 *E. coli* isolated (17/26, 65.36%) were recovered from the examined meat products in the following manner, minced meat, 5/11, 45.45% sausage (9/10, 90%), beef burger (2/3 66.66%) and basterma (0.5, 50%) were identified as *E. coli* biovar I. The other *E. coli* isolates (9.26, 34.61%) were proved to be biovar II, which recovered from minced meat (6 isolates) while in case of sausage, beef burger and basterma (one isolate each).

*Escherichia coli* in meat products.

By using multiple tube fermentation technique (MPN) *E. coli* could be detected in (30%) of the samples as compared with frequency isolation by the direct plate count technique (26%). 21 strains (70%) of 30 *E. coli* isolated from examined samples by MPN technique were proved to be *E. coli* biovar 1.

The aforementioned results revealed that the incidence of *E. coli* was high in fresh minced meat & sausage. Similar findings were recorded by El-Khatib, (1982); Gobran, (1985); Abd El-Aziz, (1987) and Niazi and Refai (1988).

From Table (2) no marked differences in the enumeration of *E. coli* in different tested meat products samples were found when the two techniques were used, a direct plate technique (DPC) and multiple tube fermentation technique (MPN). The highest mean value of *E. coli* numbers ( $4.85 \pm 0.46 \log_{10}$  (DPC),  $5.53 \pm 0.39 \log_{10}$  (MPN) per gram and ( $4.28 \pm 0.16 \log_{10}$  DPC,  $366 \pm 0.27 \log_{10}$  (MPN) per gram was recorded in minced meat beef burger respectively. The lowest value ( $2.73 \pm 0.18 \log_{10}$  (DPC),  $2.33 \pm 0.27 \log_{10}$  (MPN)/gm) was reported in basterma samples. Furthermore, statistical analysis of the enumeration of *E. coli* in tested meat products samples showed no significant difference between counts given by the two techniques. These findings confirms the previous findings reported by Andreson and Baird Parker (1975)

The results recorded in Table (3) reveal that out of 30 isolates of *E. coli* recovered from different tested meat products samples, only 12 isolates (40%) could be serologically typed as enteropathogenic *E. coli*. These strains revealed 7 different classic EPEC serovars namely O<sub>124</sub>:K<sub>12</sub> (B<sub>17</sub>) (4 strains, O<sub>44</sub>:K<sub>74</sub> (L) and O<sub>88</sub> : K<sub>61</sub> (87) (2 strains each) and one

Table (1): Incidence of E.coli (Biotype I and II) isolates among different meat products using direct plate count and Multiple tube fermentation technique

Samples	Technique	No. of examined samples	Positive samples		Escherichia coli (biotype)			
			No.	%	Biovar I		Biovar II	
			No.	%	No.	%	No.	%
Minced meat	A	25	11	44	5	45.45	6	54.54
	B	25	12	48	8	66.66	4	33.33
Suasage	A	25	10	40	9	90.0	1	10.00
	B	25	12	48	9	75.0	1	25.0
Beef burger	A	25	3	12	2	66.66	1	33.33
	B	25	4	10	3	75.0	1	25.0
Basterma	A	25	2	8	1	50.0	1	50
	B	25	2	8	1	50.0	1	50
Total	A	100	26	100	17	65.38	9	34.62
	B	100	30	100	21	70	9	30.0

A = Direct plate count technique

B = Multiple tube fermentation technique

Table (2): Statistical analysis of E.coli count (count/g  $\log_{10}$ ) in different meat products

	Minced meat		Sausage		Beefburger		Basterma	
	A	B	A	B	A	B	A	B
Minimum	3.07	2.00	2.3	2.00	3.90	3.00	2.47	2.00
Maximum	7.44	7.00	6.81	8.00	4.47	4.00	3	3.00
Mean value	8.85	5.53	3.602	3.45	4.28	3.66	2.73	2.33
S.E. $\pm$	$\pm 0.46$	$\pm 0.39$	$\pm 0.42$	$\pm 0.51$	$\pm 0.16$	$\pm 0.27$	$\pm 0.18$	$\pm 0.27$

A = Direct plate count technique

B = Multiple tube fermentation technique.

S.E. = Standard error.

Table (3): Enteropathogenic strains of E.coli (EPEC) isolated from meat product samples

Source	No. of (+)ve samples	Enteropathogenic serovars		
		O:K (B or L) serovar	No.	%
Minced meat	12	O <sub>44</sub> : K <sub>74</sub> (L)	2	16.66
		O <sub>124</sub> : K <sub>72</sub> (B <sub>17</sub> )	2	16.66
		O <sub>127</sub> : K <sub>63</sub> (B <sub>8</sub> )	1	8.33
		Untypable	7	58.33
Sausage	12	O <sub>78</sub> : K <sub>80</sub> (B-)	1	8.33
		O <sub>86</sub> : K <sub>61</sub> (B <sub>7</sub> )	2	16.66
		O <sub>111</sub> : K <sub>58</sub> (B <sub>4</sub> )	1	8.33
		O <sub>124</sub> : K <sub>72</sub> (17)	2	16.66
		Untypable	6	50.00
Beef-burger	4	O <sub>25</sub> : K <sub>11</sub> (L)	1	25.00
		Untypable	3	75.00
Basterma	2	Untypable	2	100
Total	30	Typed	12	40.00
		Untypable	18	60.00

*A.M. Darwish et al.*

strain from each of O<sub>25</sub> : K<sub>11</sub> (L), O<sub>78</sub> : K<sub>80</sub> (B-), O<sub>111</sub> : K<sub>58</sub> (B<sub>4</sub>) and O<sub>127</sub> : K<sub>63</sub> (B<sub>8</sub>)

The serovar O<sub>124</sub> : K<sub>72</sub> (B<sub>17</sub>) was recovered from fresh raw minced meat (2 strain) and fresh sausage (2 strain). The serovars O<sub>44</sub> : K<sub>74</sub> (L) (2 strain) and O<sub>127</sub> : K<sub>63</sub> (B<sub>8</sub>) (one strain) were isolated only from raw minced meat, whereas serovars O<sub>78</sub> : K<sub>80</sub> (B-), O<sub>86</sub> : K<sub>61</sub> (B<sub>7</sub>) and O<sub>111</sub> : K<sub>58</sub> (B<sub>4</sub>) were recovered from fresh raw sausage and the serovar O<sub>25</sub> : K<sub>111</sub> (L) was obtained only from the frozen beef-burger.

None of *E. coli* strains recovered from basterma slices were enteropathogenic, these findings are nearly similar to those findings recorded by Gobran (1985), Niazi and Refai (1988) for *E. coli* serotypes isolated from raw minced meat, fresh sausage and beef burger.

Most of the detectable classic enteropathogenic *E. coli* isolated from meat, sausage and beef-burger were previously reported to be incriminated in different infantile diarrhoea and gastrointestinal outbreaks in adult human (Bray, 1945; Dupont et al., 1971; Dean et al., 1972; Levine et al., 1978 and Back et al., 1980).

#### SUMMARY

One hundred samples of raw meat products, fresh minced meat, fresh sausages, frozen beefburger and basterma slices, 25 each, were collected from different markets in Cairo.

The samples were examined for determination of the incidence and enumeration of *E. coli* by using direct

*Escherichia coli* in meat products.

plate count technique (DPC) and multiple tube fermentation technique (MPN) as well as biochemical & serological identification of the enteropathogenic *E. coli*

The incidences of *E. coli* were 26% & 30% of the examined meat products by using DPC & MPN respectively.

The incidence of *E. coli* in minced meat sausage, frozen beef-burger and basterma slices was 44%, 40%, 12% and 8% by using DPC, and 48%, 48%, 16%, and 8% by using MPN respectively.

Of 30 *E. coli* isolates, 12 (40%) possessed the classic enteropathogenic *E. coli* serovars O<sub>124</sub> : K<sub>72</sub> (4 strain, O<sub>68</sub> : K<sub>61</sub>, O<sub>44</sub> : K<sub>74</sub> (2 strain each), O<sub>25</sub> : K<sub>11</sub>, O<sub>78</sub> : K<sub>80</sub>, O<sub>111</sub>, K<sub>58</sub>, O<sub>127</sub> : K<sub>63</sub> (one strain each).

Principles for production of meat products of good microbiological quality were discussed as well as public health significance of EPEC.

#### REFERENCES

1. Abd-El-Aziz, A.A. (1979): Studies on hygienic quality of manufactured fresh sausage. M. of Public Health Sci., Thesis, High Inst. Public Health, Alexandria University.
2. Anderson, Judith M. and Baird-Parker, A.C. (1975) A rapid and direct plate method for enumerating *Escherichia coli* biotype 1 in food. *J. Appl. Bact.* **39**, 111.
3. Anon, (1978): Food-borne diseases, bacteria. In international commission on Microbiological specifications for food, *Microorganisms in foods*. 1. Their significance and Enumeration, 2<sup>nd</sup> ed. P. 22. University of Toronto Press., Toronto.



*A.M. Darwish et al.*

- 4 . Agres, J.C. Mandt, J.O. and Sandine, W.E. (1980): Microbiology of foods. W.H. Freeman and Comp. San Francisco, USA.
- 5 . Back, E. Blomberg, S., Kaijeser; B., Stintzing, G. Woodstrom, T. and Habte, D. (1980): Enterotoxigenic *Escherichia* and other Gramnegativ bacteria of infantile diarrhoea, surface antigen and loss of enterotoxigenicity. *J. Infect. Dis.*, 143, 318.
- 6 . Barraud, C.; Kitchell, H.G.; Labots, H.; Rentev, G. and Simonsen, B. (1967): Standardization of the total aerobic count in meat and products. *Fleischwirtschaft*, 47, 1313.
- 7 . Bray, J. (1945): Isolation of antigenically homogenous strains of *Bact. Coli Neopolitanum* from summer diarrhoea of infants. *J. Path. and Bact.* 57. 239.
- 8 . Cruickshank, R.; Duguid, J.P.; Marmion, B.P. and Swain, R.H.A. (1975): Medical Microbiology "The Practice of Medical Microbiology. 12<sup>th</sup> Ed. Vol. II Churchill Livingstone, Edinbrough London and New-York.
- 9 . Dean, A.G.; Ching, Y.C.; Williams, R.G. and Harden, L.B. (1972): Test for *Escherichia colie*-nterotoxin using infant mice application in a study of diarrhoea *J. Infect. Dis.*, 125, 407.
10. Delepine, S. (1903): Food poisoning and epidemic diarrhoea *J.A.M.A.* 40, 657.
11. Dupont, H.L.; Formal, S.B.; Formal, S.B.; Hornick, B.B., Snyder, M.J.; Libonati, J.P., Sheehan, D.G., Labrec, E.H. and Kalas, J.P. (1971): Pathogenesis of *Escherichia coli* diarrhoea *N. Engl. J. Med.*, 285,

*Escherichia coli* in meat products.

12. Edelman, R.; and Levine, M.M. (1983): Summary of a weekshope on enteropathogenic *Escherichia coli*. J. Infect. Dis. 147, 1108.
13. El-Khatat, T. (1982): Sanitary condition of sausage in Assiut M.V.Sc., Thesis, Assiut University.
14. El-Mossalami, E. (1958): Do surface bacteria of dressed cattle increase in chilling room. Vet. Med. J. 5, 5: 113-127.
15. Frazier, W.C. and Westhoff, D.C. (1978): Food Microbiology 3rd Ed. Tata McGrow Hill Publ.Comp. Ltd, New Delhi.
16. Gobran, R.A. (1985): Enterobacterioacea in meat products in Upper Egypt. M.V.Sc. Thesis, Assiut University.
17. ICMSF. (1978): International Commission on Microbiological specification for food. Microorganisms in foods, their significance and methods of enumeration, 12th. Ed. Univ. of Toronto Press, Toronto. Buffaloe, Canada.
18. Jawetz, E., Melnick, J.L. and Adelbery, E.A. (1982): Reciew of medical Microbiology. 16th Ed. Middle East Edition.
19. Kleeberger, A.; Schafer, K. and Busse, M. (1980): Ecology of Enterobacteria on slaughter house meat. Fleisch wirtschaft, 60, 8, 1529.
20. Levine, M.M.; Berguist, E.J. Nalin, D.R.; Waterman, D.H., Hornich, R.B. Young, C.R. Stomon, S. and Rowe, B. (1978): *Escherichia coli* strains that cause diarrhoea but do not produce heat labile or heat-stable enterotoxins and are nor-invasive lancet, 1, 1119.

*A.M. Darwish et al.*

21. Matsievskii, V.; Logachev, A. Fedorina, A. and Risklova, A. (1971): Outbreak of food poisoning caused by *E. coli* O<sub>124</sub> : K<sub>72</sub> (B<sub>17</sub>). *Zhurnal Mikrobiologu, Epidemiolog. Immunobiology* 48, 137.
22. Mehlman, J.J. and Bomero, A. (1982): Enteropathogenic *E. coli*, Methods for recovery from foods. *Food Tech.*, 36, 3.
23. MIG, (1974): Microbiology laboratory Guidebook United states Department of Agriculture. Food safety and inspection service, Washington, D.C. 20250.
24. Mossel, D.A.A. and Vega, C.L. (1973): The direct enumeration of *Escherichia coli* in water using Macconkey's at 44°C in Plastic pouches. *Hlth Lab. Sci.* 10, 303.
25. Niazi, Z.M. and Refai, M. (1988): Isolation of enteropathogenic and enterotoxigenic *Escherichia coli* from meat and cheese. *Vet. Med. J.* 36,127.
26. Nickanen, A. and Pouja, M.S. (1977): Comparative studies on the sampling and investigation of Microbial contamination of surface by the contact plate and swab method. *J. Appl. Bact.* 42,53.
27. Pyathin, K. and Kriivoshein, Y. (1980): Microbiology with virology. 2<sup>nd</sup> Ed. MIR Publ., Moscow, USSR.
28. Stolle, A. (1981): Spreading & Salmonellae during cattle slaughtering. *Journal of Applied Bacteriology*, 50, 239.