

ORGANOCHLORINE PESTICIDE RESIDUES IN MEAT AND EDIBLE OFFALS

By
KHALAFALLA, F. A.*; HODA, A. AWAD** and AIDA, F. GERGIS***

* Food Hygiene Department, Fac. of Vet. Medicine, Beni-Suef, Egypt,

**Food Hygiene Department, Animal Health Res. Institute, Dokki, Egypt,

(Received : 8/12/1993)

SUMMARY

Samples were collected from 350 carcasses of slaughtered animals (175 beef, 150 buffalo and 25 mutton). Moreover, 200 samples of fresh and frozen livers were taken. With regard to chickens; skin, abdominal fat, liver, kidney and gizzard samples were collected from 50 carcasses 100 samples of fish flesh, 50 from each of *Clarias lazera* and *Tilapia nilotica*.

The collected samples were examined for organochlorine residues by using thin layer chromatoplate. The incidence and the detectable level of organochlorine residues were varied dependent on the species of animals, type of tissue examined as well as the degree of accumulation of these compounds. None of examined fat samples of slaughtered animals exceeded the permissible limits of DDT while beef muscle and fat as well as buffalo fat showed high level of lindane residue. Broiler carcasses showed no detectable level of any pesticide residues while the detectable level in laying hen carcasses did not exceed the permissible limits of aldrin and dieldrin. No detectable level of organochlorine residues in frozen liver as compared with fresh ones which contained low residual levels of aldrin, dieldrin, lindane and DDT.

Clarias lazera contained high level of dieldrin than other pesticide residues while none of detectable levels of pesticides in *Tilapia nilotica* exceeded permissible limits.

The sources of contamination to animals, chicken and fish with organochlorine pesticides were discussed.

INTRODUCTION

The widespread acceptance of chlorinated hydrocarbon was due to the fact that they are chemically stable, potent, and extremely persistent when applied to plants, food crops, or other surfaces. In the early 1960, the general public became aware that the use of these biologically active compounds were in fact, a mixed blessing. Public awareness is currently focused on the wide spread use of pesticides in the environment and their relationship to environmental health.

The problem of pesticide residues in food has been addressed at the international level through several committees sponsored by the United Nations Organisations FAO/WHO. Contamination of foods of animal origin with organochlorine pesticides and their metabolites has been reported from various countries (Neumann, 1988; Goldman et al., 1990 and Sandhu, 1992). Residues of organochlorine pesticides were reported in various foods as meats (Gergis, 1983 and Khalafalla and Gergis, 1991), poultry. (Ritchey et al., 1967 and Mc-Dougall et al., 1989) and fish (Dogheim et al., 1988; Mc-Dougall et al., 1989 and Bakre et al., 1990). The main side effect of environmental pollution by pesticides is food contamination leading to injury of nontarget organisms concerns the health of the workers and consumers. Many investigations were conducted to correlate between certain disease conditions of non etiological agents and the degree of environmental pollution with pesticide residues (Ingbedioh, 1991 and Heise, 1992).

The present investigation aimed to determine the level of contamination of some animal products

with organochlorine residues.

MATERIAL AND METHODS

I-Collection of samples:

Individual 50-100 gram samples of muscle, fat, liver and kidneys, were obtained from both cattle and buffaloes (150 each) slaughtered at Cairo and Giza as well as 25 mutton carcasses were represented by muscle, fat, liver and kidneys were examined. The investigated animals were of different age and sex. Moreover, 200 samples of fresh and frozen livers were collected from butcher shops. With regard to chickens: skin, abdominal fat, liver, kidney and gizzard samples were collected from 50 carcasses both of broilers and laying hens. The investigation concluded also the detection of level of organochlorine residues in 100 samples of fish flesh, 50 samples from each of *Claris lazera* and *Tilapia nilotica* collected from Giza markets.

The collected samples, were packed separately in sterile polyethylene bags in ice-box and examined as soon as their arrival to the laboratory.

II- Preparation of samples:

25 grams of each sample were grinded with 100 grams of anhydrous sod. sulphate, then stirred for one minute with 200 ml of petroleum ether. Solvent layer was obtained and then fat re-extracted again. Petroleum ether was evaporated to obtain extracted fat. The pesticide residues were extracted from fat by using Acetonitrile saturated with petroleum ether. (Horwitz, AOAC, 1975).

III- Calculation of pesticide residues:

Thin layer chromatoplate was used to calculate the concentration of the pesticide residue in mg/kg of samples according to technique of Horwitz, AOAC (1975)

$$\text{mg/kg} = \frac{S}{W}$$

S = Concentration of pesticide standar $\mu\text{g/ml}$

W = Weight of unknown sample.

RESULTS AND DISCUSSION

Table 1 summarizes the results of recognition of

various organochlorine pesticides in different samples. Lindane and DDT were detected in all red meat samples, while endrine was detected in buffalo samples and dieldrin in beef samples. It is worth mentioning that aldrin, dieldrin, lindane and DDT residues were frequently detected in fresh liver samples. while no detectable level of any organochlorine pesticide residues was recognised in the frozen liver samples. Moreover, the fresh liver samples were subjected for contamination with organochlorine pesticide residues and compared with liver samples collected from different carcasses at slaughter house. The data obtained during this investigation indicated that the fat and liver samples should be considered the samples of choice for recovery of pesticide residues. This agrees with that reported by Gergis (1983) and Khalafalla and Gergis (1991). This may be attributed to way of nutrition (graze in different pastures) and continuously exposure to the spraying with insecticides to control external parasites. On the other hand, the misuse of pesticides in atomised containers in butcher's shop may be responsible for the high incidence of residues in fresh liver samples.

Concerning chickens, aldrin and dieldrin residues were the only pesticides detected in laying hen samples. On the other hand, no detectable level of any pesticide was detected in broiler samples analyzed. This may be attributed to frequently spraying of laying chicken with pesticide to avoid external parasites specially in native breeds as well as pesticides particules may be drift on drinking water. Nearly similar findings were reported by Putnam et al., (1974) and Mc-Dougall et al. (1989).

With regard to fish samples; aldrin, dieldrin, and DDT residues in *Clarias lazera* showed higher frequency than in *Tilapia nilotica*. However, the highest level of the dieldrin was detected in *Claris lazera* than other meat samples. This agrees with that reported by Dogheim et al. (1988) and Mc-Dougall (1989). From the present data could be concluded that the high incidences of detection were obtained in fish samples (*Clarias lazera*), followed by laying hen samples and then liver and finally the carcasses of slaughtered animals but no detectable levels in broiler samples and frozen livers samples. The detectable level

Organochlorine pesticides residues

Table (2): Residues of organochlorine pesticides residues in various animal tissues

Sample	No. of exam. samples	Aldrin		Dieldrin		Endrin		Lindane		DDT	
		No.	%	No.	%	No.	%	No.	%	No.	%
Beef carcasses	174	4	2.3	5	2.8	2	1.1	2	1.1	2	1.1
Muscle		4	2.3	5	2.8	2	1.1	2	1.1	2	1.1
Fat		0	0	1	0.6	1	0.6	0	0	1	0.6
Liver		0	0	0	0	0	0	0	0	0	0
Kidney		0	0	0	0	0	0	0	0	0	0
Buffalo carcasses	154	0	0	0	0	0	0	0	0	0	0
Muscle		0	0	0	0	0	0	0	0	0	0
Fat		0	0	0	0	0	0	1	0.7	1	0.7
Liver		0	0	0	0	2	1.3	2	1.3	1	0.7
Kidney		0	0	0	0	1	0.7	0	0	0	0
Chicken carcasses	21	0	0	0	0	0	0	0	0	1	4
Muscle		0	0	0	0	0	0	0	0	1	4
Fat		0	0	0	0	0	0	4	16	3	12
Liver		0	0	0	0	0	0	0	0	1	4
Kidney		0	0	0	0	0	0	0	0	0	0
Truck liver	20	0	0	0	0	0	0	0	0	0	0
Truck liver	150	2	1.3	8	5.3	0	0	1	0.7	8	5.3
Truck carcasses	100	0	0	0	0	0	0	0	0	0	0
Muscle		0	0	0	0	0	0	0	0	0	0
Fat		0	0	0	0	0	0	0	0	0	0
Liver		0	0	0	0	0	0	0	0	0	0
Kidney		0	0	0	0	0	0	0	0	0	0
Carcass		0	0	0	0	0	0	0	0	0	0
Living fish carcasses	20	0	0	0	0	0	0	0	0	0	0
Muscle		4	20	5	25	0	0	0	0	0	0
Fat		0	0	0	0	0	0	0	0	0	0
Liver		4	20	2	10	0	0	0	0	0	0
Kidney		1	5	1	5	0	0	0	0	0	0
Carcass		1	5	1	5	0	0	0	0	0	0
Cattle bones	50	0	0	12	24	0	0	2	4	6	12
Truck residue	50	2	4	8	16	0	0	2	4	4	8

well-residues were varied in quantities dependent on the species of animal, type of tissue examined and exposure of examined animals to different residues before slaughter as well as the degree of accumulation of these compounds in the examined tissues (Table 2).

The maximum residual limits for DDT, lindane, Aldrin in carcass fat are 7.2 and 0.2 mg/kg (The FAO/WHO Food standard Programme, Codex Alimentarius Commission, Rome, (1978)). Most of examined fat samples of each of beef, buffalo, mutton and chicken carcasses exceeded the permissible limits of DDT residue while only one sample of buffalo liver (9.6 ppm) exceeded the limits. Concerning the lindane residue, beef muscle and fat as well as buffalo fat showed higher level of residue than other examined samples while one of dieldrin residual level in examined samples exceeded the acceptable limit. It can be concluded that the highest level of pesticide residues were reported in fats as compared

with livers, muscles and Kidneys of examined samples. This could be attributed to the fact that chlorinated hydrocarbons are fat soluble and tend to be stored more extensively in adipose tissue than other tissues. Such residues may be released from fat depots during starvation, however true excretion may occur by way of urine, faeces, milk, egg, and also crosses the placental barrier into foetal liver. This held the view reported by Ecobichon and Saschenbrecker (1968).

The detectable level of pesticide residue in livers may be attributed to the great metabolism of these substances by liver enzymes. Greater excretion in urine may be the reason for the neglectable levels of pesticide residues in the Kidneys. Feeding stuffs and the control of ectoparasites are the contributing factors for the contamination of meat with different types of pesticides. Grossklaus (1978), assumed that up to 50% of all chlorinated hydrocarbons ingested by man are derived from foods of animal origin.

Table (1): Levels of organochlorine pesticides residues in meats and edible offals

Sample	No. of exam. samples	Aldrin	Dieldrin	Endrin	Lindane	DDT
Beef carcasses	100		0	0	1.13	1.6
Muscle			0.1	14	8.8-12	1.7
Fat			0	4	0	4
Liver			0	0	0	0
Kidney						
Buffalo carcasses	100			0	0	0
Muscle				0	0	4
Fat				0	0	4
Liver				0	4	16
Kidney				0	0	0
Mutton carcasses	10					
Muscle			0		0	4
Fat			0		0.5	1.7
Liver			0		0	0
Kidney			0		0	0
Fresh liver	100	0.018-0.022	0.015-0.018	0	0.001	0.02-0.27
Frozen liver	100	0	0	0	0	0
Broiler carcasses	30	0	0	0	0	0
Skin		0	0	0	0	0
Abd. fat		0	0	0	0	0
Liver		0	0	0	0	0
Kidney		0	0	0	0	0
Gizzard						
Laying hen carcasses (one year)	30					
Skin		0.02-0.31	0.013-0.062	0	0	0
Abd. fat		0.021-0.092	0.032-0.12	0	0	0
Liver		0.003-0.052	0.013-0.05	0	0	0
Kidney		0.02	0.03	0	0	0
Gizzard		0.018	0.012	0	0	0
<i>Clarias lazera</i>	50	0.011-0.047	0.027-1.1	0	0.002-0.017	0.01-6.8
<i>Tilapia nilotica</i>	50	0.01-0.09	0.05-0.32	0	0.01-0.15	0.11-57

Broiler carcasses showed no detectable level of any pesticide residues while the detectable level in laying hen carcasses did not exceed the permissible limits of aldrin and dieldrin residues recommended by the Joint FAO/WHO (1978) (Table 2). Pesticides are used against undesirable plants and insects that injure, destroy, or cause disease to animals and humans. The promiscuous use of such compounds may be continuously exposed the animals to chemicals which accumulated in its meats and consumed by humans leading to public health hazards. Moreover, the continuous exposure of poultry feed to low level of organochlorine pesticides led to appearance of measurable residues in the abdominal fat of chickens. This held the view reported by Grossklaus (1978), the Dougall et al., (1989) and Igbedioh (1991). In this respect, Putnam et al., (1974), stated that poultry house contaminated with small amounts of pesticides from isolated pest treatments may also serve to increase the risk of tissue residues.

The detectable level of aldrin, dieldrin, lindane and DDT residues in fresh liver did not exceed the permissible limits while no detectable level in frozen livers. This may be attributed to the effect of freezing which destructed the residues. This agrees with view reported by Gergis (1987) and Khalafalla and Gergis (1991). On the other hand Lane et al., (1979) reported that the major portion of DDT was converted to DDD during storage at 18°C for 18 months.

Clarias lazera contained high level of dieldrin than other pesticide residues that exceeded the recommended limits (0.2 mg/kg) while none of the detectable levels of pesticide residues in *Tilapia nilotica* exceeded such limits. High level of pesticides residues in *Clarias lazera* can be justified by the way of nutrition and the content of the particular species of fish. *Clarias lazera* is bottom feeder, as it mostly feeds on food from the bottom of water where is considered more contaminated with chlorinated hydrocarbons.

is well as exposed to more concentration of such compounds. Nearly similar view, were reported by Enghem et al., (1988) and Bakre et al., (1990). In this respect, Mc-Dougall et al., (1989), reported that fat content and residues in the fish would be higher during the warmer season of the year.

Consequently, organochlorine pesticide residues in food of animal origin are substantially high in developing countries than in areas of intensive urban due to application of pest control programs with care under official supervision.

REFERENCES

- Mishra, P. P., Mishra, V. and Bhatnagar, P. (1990): Residues of organochlorine insecticides in fish from Mahala water reservoir, Jaipur, India. *Bull. Environ. Contam. Toxicol.* 45, 304.
- Enghem, S. M.; Almaz, M. M.; Kostandi, S. N. and Hegazy, M. E. (1988) Pesticide residues in milk and fish samples collected from Egypt *J. Assoc. Off. Anal. Chem.* 71, 674.
- Forbush, D. J. and Saschenbrecker, P. W. (1968): Study of DDT in cockerels, *Canad. J. Physiol. Pharmacol.* 46, 785.
- Gergis, A. F. (1983): Pesticide residues in meat. M. V. Sc. Thesis, Fac. Vet. Med., Cairo University, Egypt.
- Goldman, L. R., Smith, D. F.; Neutra, R. R.; Saunders, L. D.; Pond, E. M.; Stration, J.; Waller, K.; Jackson, R. J. and Kiger, K. W. (1990): Pesticide food poisoning from contaminated watermelons in California, 1985. *Arch. Environ. Health* 45, 229.
- Grossklaus, D. (1978): The occurrence and importance of residues in meat and their evaluation within official meat inspection. *Ann. Ist. Super. Santa*, 14, 319.
- Heise, S. (1992): Pesticide residues in food as a potential hazard in the third world. 3rd World Congress Foodborne infections and Intoxications. Vol. II, Berlin.
- Horwitz, W. (1975) Pesticide residues PP. 518 in 12th Ed. Official methods Analysis of the Association of Official Analytical Chemists, Washington, U. S. A.
- Ingbediob, S. O. (1991) Effects of agricultural pesticides on human, animals and higher plants in developing countries. *Archives of Environmental Health*, 45, 218.
- Joint FAO/WHO (1978): Food standard Programme, Codex Alimentarius commission, Rome.
- Khalafalla, F. A. and Gergis, A. F. (1991): Organochlorine pesticide residues in carcasses of some food animals. Beni-Suef, *Vet. Med. Res.* 3, 61.
- Lane, L. G.; Ammerman, G. R.; Lane, H. and Muir, W. M. (1979): A comparison of the influence processing and broiling on naturally occurring and spiked residues of 1,1,1-Trichloro-2,2-Bis (P-Chlorophenyl) ethane and its metabolites. *J. Agric. Food. Chem.* 27, 1156.
- Mc-Dougall, K. W.; Ahmed, N.; Harris, C. R. and Higginson, F. R. (1989): Organochlorine insecticide residues in fish and birds from three river systems on the north Coast region of New South Wales. *Bull. Environ. Contam. Toxicol.* 42, 884.
- Neumann, G. B. (1988): The occurrence and variation of organochlorine pesticide residues detected in Australian livestock at slaughter *Acta Veterinaria Scandinavica* 84, 299.
- Putnam, E. M.; Brewer, R. N. and Cottier, G. J. (1974): Low level pesticide contamination of soil and feed and its effect on broiler tissue residue. *Poultry Science* 53, 1695.
- Ritchey, S. J.; Young, R. W. and Essary, E. O. (1967): The effects of cooking on chlorinated hydrocarbon pesticide residues in chicken tissues. *J. Food Science* 32, 238.
- Sandhu, T. S. (1992): Pesticides in foods. 3rd World Congress Foodborne Infections and Intoxications, Vol. II, Berlin.