# SEROLOGICAL STUDIES ON FLOCKS SHOWING DEPRESSED EGG PRODUCTION

# NAWAL M. A. YOUSSEF and M. H. H. AHMED\*

• Dept. of Virology, Animal Health Res. Insitutute, Dokki, Giza.

## **SUMMARY**

A serological investigation was undertaken on flooks with depressed egg production. This syndrome is characterized either by a failure to attain predicted production targets or by a fali in egg numbers. The depression in production could reach over 30%. It might or might not return to normal. For a short perod the eggs produced in some flocks were smaller, lose colour, have poor egg shell strength and many soft shelled eggs are laid. Also, some flocks were suffering form mild respiratory symptoms and increase in mortality with decrease in feed intake. The results revealed that no obvious correlation was found between antibody to adenovirus, (BDS76 and CELO), infectious bronchitis virus (I.B.V), infectious larengotracheitis virus (I. L. T. V.), avain encephalomyelitis virus (A. E. V.), Newcastle disease virus (N. D. V.), reovirus or infectious bursal disease virus (I. B. D. V.) and this syndrome.

The ELISA results of turkey rhinotracheitis (TRT), showed only and positive cases out of 38 samples from four flocks, while three gave high positive results. When the same serum samples were examined by Ornithobacterium rhinotracheale (ORT) ELISA 23 of 38 serum

samples were positive to ORT and 11 serum samples ± and only 3 serum samples were negative.

The Constitution of the Co

nes extractiful and object

These results indicate that ORT might be the cause of this syndrome. This is the first serological evidence of its presence in Egypt. There was a high correlation between results of immunocomb IgG and HI for ND virus antibody titers for all serum samples tested. Immunocomb IgG proved very sensitive in detecting antibodies against IB, IBD, when it was compared with AGPT.

# INTRODUCTION

Depressed egg production occurs in broiler breeders where the syndrome affects the birds at the peak of production or soon before entering production, mostly between 24th and 52nd weeks of age (Van Eck et al., 1976).

or a relative to the second of the second of

The severity of this syndrome is extremely variable because it is influenced by many factors such as poor management, inadequate ventilation, high stocking density, poor litter condition, poor hygiene, high ammonial level and concurrent disease. Debeaking or vaccination with live

Newcastle disease virus if done at a critical time (Hafez, 1994).

During viruological investigation of these outbreaks a number of agents were recognized as egg drop syndrome 076 (Khafagy and Hamouda, 1991). EDS Giza 1991 (Ahmed, 1995), turkey rhinotracheitis (TRT), and other microorganisms were designated as Pasteurellia-like organism (ORT). Vandamme et al., (1994) carried out further identifications on this microorganism using genetic taxonomic methods and designated that bacterium as Ornithobacterium rhinotracheale gen, nov sp. (nov. in the rRNA-superfamily V).

# The purposes of the present study were

- 1- Detection of specific antibodies to some common viruses that have been incriminated as causes of depressed egg production.
- 2-Evaluation of the application of the immunocomb bursal disease - Newcastle -Bronchitis antibody test kit as a quick sensitive test which detects antibody levels in the serum and compare its results with the results of HI and AGPT.

#### MATERIAL AND METHODS

#### Serum samples:

Blood was collected from broiler breeders (30 t 51 weeks old), their details are shown in Table (1). These broiler breeders were in flocks vaccinated against. Marek's disease, infectious bronchitis, Newcastle disease, Reodisease, infectious aryngotracheitis, infectious bursal disease and

avian encephalomyelitis. The blood was refrigerated at 4°C overnight, the serum was then separated by centrifugation at 1500 rpm for 10 mins, the clear serum was then collected and kept in the deep freeze at -20°C.

Haemagglutination (HA) and haemagglutination inhibition (HI) tests: The tests were conducted by microtiter method of ND acording to Beord and Wilkes (1973) and for EDS76 according to Mcferran (1989).

Agar gel diffusion tests (AGDT): The test was carried out according to the method of Hitchner et al., (1975) to detect the antibodies against IB, IBD, AE and Celo virus. Positive and negative sera included in the test were kingly supplied by (Doorn Institute, Holland)

Immunocomb solid-phase imunoassay kit: manufactured by commercial Biogal galed Labs. (Israel). This kit is designated to determine chicken serum IgG antibody titers to bursal disease-Newcastle-bronchitis (Rivetz et al., 1985). Results of immunocomb were compared with HI titer for NDV and AGD for IB and IBD viruses.

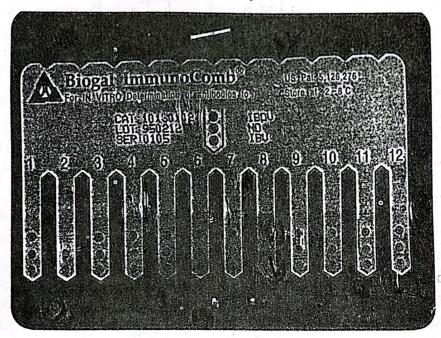
Indirect ELISA test: The test was carried out according to Hafez and Sting (1995). Serum samples from 38 broiler breeders suffering from depressed egg production and respiratory symptoms were sent to State Veterinary Laboratory. Stuttgart, Germany. Serum samples were examined for the presence of antibodies against TRT virus, and antibodies to ORT bacterium using self made indirect ELISA

prepared from Stuttgart ORT-turkey-isolate GGD 1269/91.

### RESULTS

Clinical signs: As shown in Table (1), the reduction percentage in gee number varied from 10-38. In broiler breeders the disease affects the

birds at the peak of production or soon before entering production, mostly between 30th weeks of age. Before the main symptoms are detected a slight increase in mortality in some flock's and decrease in feed intake may be observed plus the respiratory manifestation. The symptoms are generally accompanied with a drop in egg production, decrease in egg size and poor shell quality, fertility and hatchability are affected in many cases.



- \* 1 = + ve \*\* 2 = - ve
- Fig. (1): The immunocomb is a plastic card shaped like a comb on which purified IBDV, ND and IVB antigens are attached. This kit is designed to determine chick serum IgG antibody titers to these viruses. The upper spot on the immunocomb tests for IBDV, the second spot tests for NDV and the lower spot for IBV.
  - \* To determine its titer, compare the specimens colour intensity with that of the positive control (+) included in the kit and comparing with scale score.
  - \* Specimens with an identical or higher colour intensity than the positive control are considered positive.
  - \* The negative control consists of non immunized sera and should be read as zero (S-0).

resconditity and fertility were greatly reduced

		House		Flock			Flor				
200	1					7		Koss		-	
1	2900		,,,,,	7700		,000		3900	сарасну	House	
Water Le	Closed	- Jack	Closed	system		Closed	System		Type	type	
	Local		Local			Local		Local	chicken	Source of	
	38		23			30	100000	38	fall (weeks)	Age at	
	30-62		0-38			010		30-62	egg fall%	Average	
1	32		38			10		33		Maximum	
	13		12			5	10	Jall (Weeks)	maximum	7	
								production	maximum		
•	shell	less, shell-less loose					•		Egg quality		
•		•			Facial	oedema	Facial		Sings		
•		tion in the	the heart	Thage on	Petochial haemo-		•		PM		
		high			High	E TO	•		Mortality		
									Remarks		

Table (1): History of examined flocks for drop in egg production.

722

Vet.Med.J., Giza. Vol. 44, No. 4(1996)

serological results: Sera were tested for antibody to ND and EDS 76 as shown in Table (2). HI titer in Log2 ranged from 8 to 10 for ND and all sera showed negative results by HI for EDS 76 disease. The detection of antibodies for AE, Reo, ILT and adenovirus by AGD test showed that most of the sera were positive. The measurable antibodies of all viruses showed the normal levels according to the vaccination (Table 3).

Of as It has an accommod to sair of

Table (4) showed that the comb score increased with the increase in HI titer for ND. Some samples gave negative results to IB and IBD with AGDT, while gave positive results with immunocomb kit..

to tool others stook above (iii) the

Table (5) showed clearly that antibodies to ORT were detected by ELISA in 23 sera sampled from broiler breeder flocks (60.5%) and antibodies to

TRT were detected in 6 samples (15%).

Table (2): Results of tested serum by haemagglutination inhibition test (HI).

1.1	Age (weeks)	No. of	Virus	Log <sub>2</sub> HI titers \(\tau\)								Contract of			
					samples	4 . (	1	21	3 3	4	5	6	17	8	9
Hous 1	51	20	ND EDS	aic.	-			16.4				1	19		
Flock 4	30	25	ND EDS	o o o o o o o o o o o o o o o o o o o	-								25		
Flock 3	38	17	ND EDS	1.1	-		2.1	711	0	1		2	15		
house 4 +	51	15	ND EDS	1.0	-		2		C.	1	9		15		

Table (3): Detection of antibodies to IB, ILT, IBD, Adeno, AE and Reo viruses by AGDT.

31.46	Age	No. of	No. of positive serum samples by AGDT							
	(weeks)	samples	IB	ILT	IBD	Adeno	AE	Reo		
House 4	51	20	12	10	14	-ve	7.8	10		
Flock 4	30	25	15	1311	29/413	400	8.14	12		
Flock 3	38	17	10	6	10	2	8	6		
House 3	- 51	15	10	8 -	- 8	4	- 5	10		

Vet.Med.J., Giza. Vol. 44, No. 4(1996) A. Land 10-V

Table (4): Comparative study between results of immunocomb and HI to ND and

Flocks	No. of	HI titer	Immunoc	omb scale (	AGDT		
11000	samples	ND	ND	IBD	IB	IBD	IB
House 4	Samples	1024	S 5	S 6	S 4	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	+
House 4	,		5	6	3	di prijeton t	111
	1 1	46.	6	5	3	Sec. of Sec. and A	
	1 4 1	250 277734	6	5	4	+	ACADOMA A
	;		5	6	4	DA tut ex	1 + 1
	6		5	6	4	- '	+
e tro Piesero.	7	Simbon Si	5	6	4	rax grade clearan	AND BY
			Annual Control		3	cornect restrict	hi ni y
ALTER HITTER	8 9	3 1 500	5	5	3	+	-
less from I	10		6 5 6	5	4	State State of the	4
Flock 4	10		5	5	3	•	+
1004 4	The state of the s			6	3	+	+
	2 3		0	6	3 3 4	+	
			6 6 5 6 6	6 5 5 6 6 5 5 5 5 6		<u> </u>	
	4		6	3	5	(2) N - (2)	MC(2):
	4 5 6 7 8 9	in blanch	5	3	4 5 5 3 3 4		
e and the	6		6	5	3	+ 4	+
A. A.	7		6	5	3		+
61	8			6	3	+ + + +	70 E
	9		6	5	3		i de la companya de
and the last	10		6 5 5 5 6	5			
lock 3			5	6	4	- - - - +	+
1.0	1 2 3		5	6	4,	-	1-1
1.211	3		6	6	41-	++	-
				5			1.0
	4 5 6		6 6 6 5 5	5	3	+	
As A	6		0	5	5	+ 100	+
	7		0	5	5		
· L. Bild	6		)	3	3	+	+
	8		5	6	3	+	•
	9 10	De la company	6	6	3	+	+
louse 5	10	Secretary and	6	6 5 5 5 5 6 6 6 5	3 3 5 5 5 3 4 3		+
iouse 5	1		5	6	3	+	
are which	000 <b>2</b> GH	Matri, 0,336.				er anceronda	1610
W	3		6	6	3 3 4	+	- 74
	4		6	6	3	-	+
part of the	5	marin mention	5	1 6	4	1-1-14	+
d	6	agi sona a	6	6	3	+ 048	
C. C. C.	7	And the same	5	. 6	3	to de level	
1.	8	and a	6 6 5 6 5 5	6 6 6	3		
	9		6	6	3	+ 18	
	7	the second of th					

Table (5): Examination of serum samples from broiler breeder flocks

Flocks	No. of		TRT-ELIS	A	ORT-ELISA			
	samples	+	±	-	+	±		
House 4	10	3	0	7	10	0	0	
Flock 4	15	0	2	13	4	8	3	
Flock 3	5	3	1	1	2	2	1	
House 5	8	0	0	8	7	1	0	
Total	38	6	3	20	23	11	4	

props in egg production cause heavy economic losses in the poultry industry. This syndrome is accompanied with reduction of egg shell quality, decreased fertility, decreased hatchability and increased mortality rates. The serological examination of all serum samples by different tests showed that the immune response of broiler breeders in all flocks was in the normal level as a result of vaccination (Tables 2 and 3). In the investigated broiler breeders the disease primarily affected the birds at the peak of production or before entering production between 30 and 52nd weeks of age. Before the drop of egg production, a slight increase in mortality and decrease in feed intake, mild respiratory symptoms followed by facial oedema (Table 1). The results of indirect ELISA showed that the broiler breeders with antibody to ORT were 100% in house 4, 87.5% in house 5, 26, 7% in flock 4 and 40% in flock 3. The antibody percentages to TRT were 30% in house 4, 60% in flock 3. This high positive results to ORT and the presence of antibody to TRT indicate that these flocks are exposed to natural infection to ORT and TRT because there is no vaccination policy adopted in Egypt against ORT and TRT. This means that the main cause of depressed egg production in the investigated flocks was ORT. Diagnosis of ORT based on clinical signs and pathological lesions is very difficult since many other infectious diseases such as pasteurellosis, chlamydiosis, can produce similar clinical signs and postmortem lesions. Diagnosis can only be confirmed by isolation and identification of the causative agent and detection of the specific Ab against it. The epidemiology of

at the manual May 9 I multing

ORT has been discussed previously by Hafez (1994) who attributed the disease to mixed infection with Pasteurella-like organism. Vandamme et al., (1994) carried out further identification of isolates using genetic taxonomic mehtods and designated that bacterium as Ornithobacterium rhinotracheale gen, nov. in the rRNA-superfamily V. 22 12 2001

here and his guidence putting this work on the

The present study offers an excellent opportunity for a comparative investigation between the different serological tests, concerning their sensitivity and validity as diagnostic tools. Thus (4) shows as trial where the results of the different serological tests have been compared in relation to s valus of the immunocomb kits (the intensity of colour). The results of the tests showed that the HI titer of ND was parallel to S value of the immunocomb. On the other hand the low sensitivity of AGPT may explain the negative results to IBD and IBouthis is attributed to the antibody response did not reach levels sufficiently high to give positive reaction. All samples were positive to IB and IBD when tested by the immunocomb a matter which denotes clearly the superiority of HI and the immunocomb over the AGPT. The immunocomb is a self-contained portable kit. A sensitive test which detects antibody levels in the blood, serum or egg yolk, the immunocomb provides results within 40 minutes.

In conclusion, the presence of considerable levels of antibodies against ORT and TRT virus must receive more attention for deeper and planned investigation to prove the isolateion and identification of the microorganism under field

Vet.Med.J., Giza. Vol. 44, No. 4(1996)

conditions and to determine the role of these agents in the production of egg as well as its interaction with other microorganisms.

Acknowledgement: We wish to express our appreciation to Dr. Habil H. M. Hafez, Head of Poultry and Virology Department, State Veterinary Laboratory. Stuttgart, Germany, for his help and his guidance putting this work on the right track.

## REFERENCES

- Ahmed, M. H. H. (195): Viruses with drop in egg production. Ph. D. V. Sc. Thesis, Fac. Vet. Med., Cairo Univ.
- Beord, C. W. and Wilkes (1973): A simple and rapid microtest procedure for determining Newcstle haemaggiutination-inhibition (HI) antiody titers. Froc. 77 Ann. Mig. U. S. An. Health.
- Hafez, H. M. (1994): Respiratory disease condition in meat turkey caused by Ornithobacterium rhinotracheale, clinical signs, diagnostic and therapy. Proceedings of Western Poultry Disease Conference, Sacramento, P. 113-114.
- Hafez, H. M. and Sting, A. (1995): Serological surveillance on Ornithobacterium rhinotracheale in poultry flocks using self made ELISA (In preparation).

- Hitcher, S. B.; Domermuth, C. H.; Purschose, H. G. and Williams, J. E. (1975): Isolation and Identification of Avian Pathogens. American Association of Avian Pathologists. Arnold Printing Corp., Ithaca, New York, 14850.
- Khafagy, A. K. and Hamouda, M. S. M. (1991): Studies on egg drop syndrome, existence of antibodies in Egyptian commercial chicken layers. Vet. Med. J. Giza, 39 (3): 1110-1121.
- McFerran, I. B. (1989): Adenovirus: In a Laboratory Manual for the Isolation and Identification of Avian Pathogens, third edition, ASAP, Kindall Hunt Publishing Company, pp: 77-84.
- Rivetz, B. Y.; Welsman, M.; Ritterbad, F. and Hesberg, M. (1985): Evaluation of a novel rapid kit for the viral detection of Newcastle disease virus antibodies. Avian Dis., 29: 929-942.
- Van eck, J. H. H.; Davelaar, F. G.; Ven den Heuvel; Plesman, T. A. M. and Van Kol, N. (1976): Dropped production, soft shelled and shell-less egg associated with appearance of precipitation to adenovirus in flocks of laying fowl. Avian Patholoagy, 5: 261-271.
- Vandamme, P.; Segers, P.; Vancanneyt, M.; Van Hover, K.; Mutters, R.; Hommez, J.; Dewhirst, F.; Paser, K.; Derters, E.; Falsen, L; Dewroeze, M.; Bisgarrd, K.; Hinz, H. and Mannheim, W. (1994): Description of Ornitiobacterium rhinotracheale gen nov. sp. nov. isolated from the avian respiratory tract. International Journal of Systematic Bacteriology, 44: 24-37.