

## BACTERIAL ISOLATES FROM THE RESPIRATORY TRACT OF CATTLE AND THEIR ZONOTIC IMPORTANCE IN AL-QASSEEM, SAUDI ARABIA

BY

E. Y. EL-HAENAEY, O. M. MAHMOUD and E. M. HAROUN

Department of Veterinary Medicine, King Saud University, P. O. Box 1482 Buraidah, Saudi Arabia.

Received:15/12/1993

### SUMMARY

A two years survey (1990-1991) was carried out to determine bacterial respiratory pathogens of cattle. A total of 350 samples collected from the respiratory system of cattle, of which 200 from the slaughterhouse, 50 from apparently healthy cattle, 50 from stressed animals and 50 from animals with respiratory affections. Examination of these samples resulted in isolation of 123%, 134%, 186% and 182% respectively. The results showed that 325 animals were positive with an incidence of 92.9%. Of those 194 cases (59.7%) showed mixed infection and 131 cases (40.3%) had one bacterial isolate in a pure form in culture. The total numbers of were 497 (142%) of which, 137 Staphylococcus, 32 Streptococcus, 74 *E. coli*, 39 Proteus spp., 31 Corynebacterium spp., 57 anthracoides, 18 Pasteurella spp., 24 Klebsiella spp., 14 Citobacter spp., 10 Serratia spp. 9 Enterobacter spp. and 16 Pseudomonas spp. Bacteriologic examination of 150 nasal swabs from human beings that were in contact with these animals (abattoirs and farms) resulted in 25 bacterial isolates. These were 8 Staph., 4 strep., 11 anthracoides and 2 Klebsiella. The pathogenic strains were *Staph. aureus* (3), *Strep. Pyogenes* (1) and Klebsiella (2). The non-pathogenic isolates (12.7%) were *Staph. albus* (3), *Staph. citrus* (2), *Strep. fecalis* (3) and anthracoides (11).

### INTRODUCTION

*C. pyogenes*, *P. haemolytica*, *E. coli*, *Ps. aeruginosa* Haemophilus spp. and Salmonella spp. were incriminated in the etiology of respiratory diseases in cattle (Fischer, 1975; Garoiu, et al., 1982; Singh and Singh, 1980). El-Enbaawy

(1986) isolated six strains of Coryn. spp. from 137 slaughtered cattle with an incidence of 4.5%. She also sampled 159 cattle and isolated 28 strains of Coryn. spp. with an incidence rate of 16.7%. 17 strains of *P. spp.* were isolated from cattle lungs and nasal swabs and differentiated into 13 *p. multocida* and 4 *P. haemolytica*. She also obtained 113 isolates of Staph. spp. (85 Staph. epidermidis, 28 Staph aureus), 46 Strep., 22 *E. coli* and 7 strains of *Pseud. aeruginosa*. Collins et al., (1988) examined 33 diseased cattle lungs and the infection was attributed to Mycoplasma, *C. pyogenes* and *Staph. aureus*.

*Staph. aureus*, *K. pneumoniae* and *Strep. pyogenes* were isolated from the upper respiratory tract and hands of humans in contact with the animals (Galloto et al, 1988; Abd-Alla, 1990 and Saleh et al, 1993). This study was done to show the predominant respiratory bacteria in cattle and in associated human workers.

### MATERIAL AND METHODS

A total of 500 samples were collected from the respiratory system of cattle. Out of these samples were from slaughtered cattle, 150 from farm cattle and 150 from humans working in contact with investigated animals. Samples collected from cattle were nasal swabs, tracheal swabs, laryngo-tracheal swabs and heart blood. Samples were collected by sterile cotton swabs from freshly slaughtered cattle and immediately transferred to the laboratory for cultivation. Lung samples were collected from congested lung areas of the slaughtered cattle in sterile plastic bags and transported for bacteriological examination.

Collected samples were divided into two portions.



The first portion was inoculated onto nutrient agar, blood agar, mannitol salt agar and MacConkey agar plates and incubated aerobically at 37°C for 24-48 hours. Selenite "F" broth was inoculated with the second portion and incubated at 37°C for 12 hours. A loopfull was subcultured onto Hektoen enteric agar and MacConkey's medium. The inoculated plates were incubated at 37°C for 24-48 hours.

Suspected growing colonies were characterized on the basis of morphological and colonial appearance according to Finegold and Martin (1982). The pure colonies were identified biochemically according to Koneman et al (1983) and Krelg and Holt (1984). The pathogenicity and virulence of *P. multocida* to mice were determined according to Wilson and Miles (1975). Susceptibility of the most predominant pathogenic isolates to different chemotherapeutic agents was tested by the disc diffusion method according to Finegold and Martin (1982).

**RESULTS**

Table (1) shows the results of bacterial isolation from the lungs. 325 (out of 350 samples) bacterial isolates were obtained representing 92.9% of total samples investigated. Of those 194 isolates were of mixed infection and 131 were obtained in pure form (Table 2 and 3). The total isolation rate was 142% where 497 isolates were obtained. These isolates were : 173 *Staphylococcus*, 32

*Streptococcus*, 74 *E. coli*, 30 *Proteus*, *Corynebacterium*, 57 *anthracoides*, 9 *P. multocida*, 2 *P. haemolytica*, 24 *Klebsiella spp.*, *Citrobacter spp.*, 10 *Serratia spp.*, 9 *Enterobacter spp.* and 16 *Pseudomonas spp.* (Tables 4-7).

Bacteriological examination of nasal swabs from 150 humans working in cattle farms and abattoirs resulted in 25 bacterial isolates. Which typed as *Staphylococcus*, 4 *Streptococcus 11 anthracoides* and 2 *Klebsiella spp.* The pathogenic strains had an incidence of 4% and they were 3 *Staph. aureus*, 1 *Strep. pyogenes* and 2 *Klebsiella spp.* The non-pathogenic isolates were 3 *Staph. albus*, *Staph. citreus*, 3 *Strep. fecalis* and 11 *anthracoides*. All isolates from humans were identical to those recovered from cattle.

Table (B) shows the results of the susceptibility of the isolates to antibacterial agents. *Pasteurella multocida* isolates were highly resistant to ampicillin, tetracycline, cephalothin and flumequine. They varied in their sensitivity to the remaining tested drugs. Strains of *Ps. aeruginosa* were resistant to all antibiotics tested. Most strains of *C. Pyogenes* were highly sensitive to gentamicin (100% sensitivity), flumequine (91.7%) and erythromycin (50%).

**DISCUSSION**

Commensals present in the respiratory system may cause disease when animals are subjected

Table 1: Rate of bacterial isolation from respiratory system of cattle.

Source and type of samples	Number of samples	Positive		Number of isolates	Isolation rates %
		No.	Percentage		
Slaughtered cattle (Different parts of resp. system).	200	180	90%	246	123%
Living cattle (nasal swabs)	150	145	96.7%	251	167.3%
Total	350	325	92.9%	497	142%



Table (2): Types of infection and incidence of bacteriological positive cases in Slaughtered cattle.

Sources of samples	Total no. of samples	Positive samples		Positive samples		One isolate		Mixed	
		No.	No.	No.	No.	No.	No.	No.	No.
Laryngo-tracheal swabs	47	40	85.1%	7	14.9%	13	27.7%	27	57.4%
Tracheal swabs	40	36	90%	4	10%	23	57.5%	13	32.5%
lung tissues	113	104	92%	9	7.9%	47	41.6%	57	50.4%
<b>Total</b>	<b>200</b>	<b>180</b>	<b>90%</b>	<b>20</b>	<b>10%</b>	<b>83</b>	<b>41.5%</b>	<b>97</b>	<b>48.5%</b>

N.B. No. of isolates 246 (123%) .

Table (3): Results of bacteriological examination of diseased or stressed cattle.

Sources of samples	Total no. of samples	Positive samples		Positive samples		One isolate		Mixed		Total	
		No.	%	No.	%	No.	%	No.	%	No. isolates	Isolation rate%
Apparently healthy	50	47	94%	3	6%	27	54%	20	40%	67	134%
Stressed	50	48	96%	2	4%	13	26%	35	70%	93	186%
Diseased	50	50	100%	--	--	8	16%	42	84%	91	182%
<b>Total</b>	<b>150</b>	<b>145</b>	<b>96.7%</b>	<b>5</b>	<b>3.3%</b>	<b>48</b>	<b>32%</b>	<b>97</b>	<b>64.7%</b>	<b>251</b>	<b>167%</b>

Table (4): The incidence of pathogenic in samples from examined cattle

Bacterial species	Slaughtered (200)		Living								Total (350)	
			Healthy (50)		Stressed (50)		Sneezing and coughing (50)		Total (150)		No. of isolates	Isolation rate%
	No.	%	No.	%	No.	%	No.	%	No.	%		
<i>Staph. aureus</i>	40	20%	--	--	10	20%	7	14%	17	11.3%	57	16.3%
<i>Strep. pyogenes</i>	2	1%	--	--	2	4%	--	--	2	1.3%	4	1.1%
<i>C. pyogenes</i>	1	0.5%	--	--	4	8%	7	14%	11	7.3%	12	3.4%
<i>P. multocida</i>	4	2%	--	--	3	6%	6	12%	9	6%	13	3.7%
<i>P. haemolytica</i>	3	1.5%	--	--	2	4%	--	--	2	1.3%	5	1.4%
<i>Pa. aeruginosa</i>	--	--	--	--	--	--	16	32%	16	10.7%	16	4.6%
<i>Klebsiella spp.</i>	24	12%	--	--	--	--	--	--	--	--	24	6.9%
<i>E-coli</i>	40	20%	--	--	11	22%	23	46%	34	22.7%	74	21.1%
Pathogenic bacteria	114	57%	--	--	32	64%	59	118%	91	60.7%	205	58.6%
Saprophytic	132	66%	67	134%	61	122%	32	64%	160	106.7%	292	83.4%



Table (3): Incidence of Gram positive bacteria in examined cattle.

Bacterial species	Slaughtered (200)		Living								Total (350)	
			Healthy (50)		Stressed (50)		Sneezing and coughing (50)		Total (150)		No. of isolates	Isolation rate
	No.	%	No.	%	No.	%	No.	%	No.	%		
<i>Staph aureus</i>	40	20%	-	-	10	20%	7	14%	17	11.3%	57	16.3%
<i>Staph epidermidis</i>	67	33.5%	12	24%	19	38%	18	36%	49	32.7%	116	33.1%
Total	107	53.5%	12	24%	29	58%	25	50%	66	44%	173	49.4%
<i>Staph. faecalis</i>	12	6%	-	-	5	10%	5	10%	10	6.7%	22	6.3%
<i>Staph. faecalis</i>	-	-	-	-	2	4%	4	8%	6	4%	6	1.7%
<i>Staph. pyogenes</i>	2	1%	-	-	2	4%	-	-	2	1.3%	4	1.1%
Total	14	7%	-	-	9	18%	9	18%	18	12%	32	9.1%
<i>C. bras</i>	2	1%	-	-	3	6%	3	6%	6	4%	8	2.3%
<i>C. pyogenes</i>	1	0.5%	-	-	4	8%	7	14%	11	7.3%	12	3.4%
<i>C. sp.</i>	3	1.5%	-	-	6	12%	2	4%	8	5.3%	11	3.1%
Total	6	3%	-	-	13	26%	12	24%	25	16.7%	31	8.9%

Table (4): Incidence of Gram negative bacteria in examined cattle.

Bacterial species	Slaughtered (200)		Living								Total (350)	
			Healthy (50)		Stressed (50)		Sneezing and coughing (50)		Total (150)		No. of isolates	Isolation rate
	No.	%	No.	%	No.	%	No.	%	No.	%		
<i>P. multocida</i> *	4	2%	-	-	3	6%	6	12%	9	6%	13	3.7%
<i>P. haemolyticus</i> *	3	1.5%	-	-	2	4%	-	-	2	1.3%	5	1.4%
<i>Ps. aeruginosa</i> *	-	-	-	-	-	-	16	32%	16	10.7%	16	4.6%
Total	7	3.5%	-	-	5	10%	22	44%	27	18%	34	9.7%
<i>E. coli</i> **	40	20%	-	-	11	22%	23	46%	34	22.7%	74	21.1%
<i>Enterobacter</i> spp.**	4	2%	-	-	5	10%	-	-	5	3.3%	9	2.6%
<i>Citrobacter</i> spp.**	3	1.5%	2	4%	6	12%	3	6%	11	7.3%	14	4%
<i>Klebsiella</i> spp.**	24	12%	-	-	-	-	-	-	-	-	24	6.9%
<i>Frutos</i> spp.**	14	7%	3	6%	11	24%	8	16%	23	15.3%	39	11.1%
<i>Serratia</i> spp.**	2	1%	1	2%	3	6%	4	8%	8	5.3%	10	2.9%
Total	89	44.5%	6	12%	37	74%	36	76%	81	54%	170	48.6%

\* Oxidase positive  
\*\* Oxidase negative

Table (7): The total isolates of bacterial groups in examined cattle.

Bacterial groups	Slaughtered (200)		Living (150)		Total (350)	
	No.	Percentage	No.	Percentage	No.	Percentage
Staphylococci*	107	53.5%	66	44%	173	49.4%
Streptococci*	14	7%	18	12%	32	9.1%
Corynebacterium*	6	3%	25	16.7%	31	8.9%
Anthracooides*	24	12%	33	22%	57	16.3%
Total	151	75.5%	142	94.7%	293	83.8%
Serratia spp.**	2	1%	8	5.3%	10	2.9%
E-coli**	40	20%	34	22.7%	74	21.1%
Enterobacter**	4	2%	5	3.3%	9	2.6%
Pseudomonas**	--	--	16	10.7%	16	4.6%
Klebsiella**	24	12%	--	--	24	6.9%
Proteus spp.**	16	8%	23	15.3%	39	11.1%
Citrobacter**	3	1.5%	11	7.3%	14	4%
Pasteurella**	7	3.5%	11	7.3%	18	5.1%
Total	96	48%	108	72%	204	58.3%

\* GramPositive group.

\*\* Gram negative group.



Table (8): Results of antibacterial sensitivity test to some pathogenic representative bacterial isolates

Chemotherapeutic disc	Staph. aureus (20)		Staph. pyogenes (4)		C. pyogenes (12)			P. multocida (12)			P. haemolytic (5)		Ps. aeruginosa (15)		Kleb. ssp. (10)		E. coli (50)		
	R	S	R	S	R	I	S	R	I	S	R	S	R	S	R	S	R	I	S
Ampicillin	12 (60)	8 (40)	3 (75)	1 (25)	7 (58.3)	1 (8.3)	4 (33.3)	12 (100)	0 (--)	0 (--)	0 (--)	5 (100)	15 (100)	0 (--)	10 (100)	0 (--)	22 (73.3)	8 (26.7)	0 (--)
Erythromycin	10 (50)	10 (50)	4 (100)	0 (--)	3 (25)	3 (25)	6 (50)	6 (50)	1 (8.3)	5 (41.7)	5 (100)	0 (--)	10 (66.7)	5 (33.3)	9 (90)	1 (10)	21 (70)	6 (20)	3 (10)
Tetracyclin	8 (40)	12 (60)	2 (50)	2 (50)	9 (75)	1 (8.3)	2 (16.7)	10 (83.3)	0 (--)	2 (10.7)	3 (60)	2 (40)	14 (93.3)	1 (6.7)	4 (40)	6 (60)	22 (73.3)	3 (10)	5 (16.7)
Gentamicin	2 (10)	18 (90)	0 (--)	4 (100)	0 (--)	0 (--)	12 (100)	1 (8.3)	3 (25)	8 (66.7)	5 (100)	2 (40)	10 (66.7)	5 (33.3)	2 (20)	8 (80)	10 (33.3)	8 (26.7)	12 (40)
Streptomycin	14 (70)	6 (30)	3 (75)	1 (25)	8 (66.7)	0 (--)	4 (33.3)	6 (50)	3 (25)	3 (25)	4 (80)	0 (--)	8 (53.3)	7 (46.7)	0 (--)	10 (100)	19 (63.3)	0 (--)	11 (36.7)
Chloramphenicol	8 (40)	12 (60)	2 (50)	2 (25)	10 (83.3)	1 (8.3)	1 (8.3)	6 (50)	0 (--)	6 (50)	4 (80)	1 (20)	15 (100)	0 (--)	3 (30)	7 (70)	10 (33.3)	2 (6.7)	18 (60)
Cephalothin	6 (30)	14 (70)	1 (25)	3 (75)	4 (33.3)	0 (--)	8 (66.7)	12 (100)	0 (--)	0 (--)	4 (80)	1 (20)	15 (100)	0 (--)	10 (100)	0 (--)	25 (83.3)	3 (10)	2 (6.7)
Halidixic acid	8 (40)	12 (60)	3 (75)	1 (25)	2 (16.7)	4 (33.3)	6 (50)	7 (58.3)	0 (--)	5 (41.7)	3 (60)	2 (40)	9 (60)	6 (40)	1 (10)	9 (90)	20 (66.7)	0 (--)	10 (33.3)
Flumequine	10 (50)	10 (50)	2 (50)	2 (50)	0 (--)	1 (8.3)	11 (91.7)	11 (91.7)	1 (8.3)	0 (--)	5 (100)	0 (--)	8 (53.3)	7 (46.7)	2 (20)	8 (80)	20 (66.7)	5 (16.7)	5 (16.7)
Trimethoprim-sulfamethoxazole	0 (--)	20 (100)	2 (50)	2 (50)	8 (66.7)	1 (8.3)	3 (25)	7 (58.3)	0 (--)	5 (41.7)	4 (80)	1 (20)	14 (93.3)	1 (6.7)	5 (50)	5 (50)	14 (46.7)	9 (30)	7 (23.3)

Figures between parenthesis represents percentage value  
 R : Resistant.  
 I : Intermediate.  
 S : Sensitive.



stress factors (Palotay, and Newhall, 1958). Examination of 350 lung samples from cattle and 92.7% of them were harboured pathogenic bacteria. Such high incidence of isolation was also reported by El-Yas (1982) and El-Enbaawy (1986). High percentage of mixed cultures were obtained from diseased and stressed cattle. The incidence of isolation of one organism from healthy, slaughtered, stressed and diseased cattle was 54%, 41.5, 26% and 16% respectively. Similar results were reported by El-Yas (1982), El-Enbaawy (1986) Abd-Alla (1990) and Abdel Ghani et al., (1990). Higher incidence of isolation was obtained from diseased and stressed cattle as compared to slaughtered and healthy ones. The incidence of isolation of Gram positive cocci and rods was 83.8% and that of the Gram negative rods was 58.3%. Those results are similar to those previously reported by El-Enbaawy (1986).

Isolation of *E. coli* showed variable incidence rates. *E. coli* was not isolated from healthy cattle (Table 6). Magwood et al., (1969) reported that *E. coli* was seldom isolated from healthy and pneumonic herds. However, Hamdy and Trapp (1967) isolated *E. coli* in an incidence of 30.7% from normal calves before weaning and Al-Yas (1982) isolated *E. coli* in an incidence of 6% from nasal swabs of normal animals.

Pathogenic (coagulase positive) *Staph. aureus* isolates were found in 14% of the diseased cattle with mixed infection. Similar results were reported by Hamdy and Trapp (1967) and El-Yas (1982) who reported that isolation rate of 70% from nostril swabs of healthy cattle.

Streptococci were isolated from stressed and diseased animals. Hoerlin (1961), Hamdy and Trapp (1967) and Magwood et al., (1969) recovered haemolytic Streptococci at an incidence of 35.4% from the nasal swabs of calves suffering from shipping fever. *P. multocida* was isolated with an incidence of 2% from congested lungs of slaughtered cattle. *C. Pyogenes* was isolated from stressed and slaughtered cattle which confirms the previous reports of El-Enbaawy (1986) and Abdel-Ghani et al (1990). *Pseudomonas* was isolated from diseased cattle only which indicates that it is one of the most pathogenic respiratory organism (Prasad et al, 1967 and Fischer, 1975).

*Klebsiella spp.* were isolated from slaughtered cattle. El-Ebaawy (1986) obtained 14 isolates of *Klebsiella* with an incidence of 10.5% from slaughtered cattle.

Staphylococci, Streptococci, *Klebsiella* and anthracoides were isolated from humans working with animals. Similar results were reported by Lundberg et al (1984), Niemela et al (1985) Galoto et al (1988) and Saleh et al (1993).

The results have shown that *P. multocida* and *Pseudomonas spp.* were the most sensitive organisms to antibiotics as compared with *C. pyogenes* which was highly sensitive to gentamicin (100%), flumequine (91.7%) and erythromycin (50%). Wernicki and Rzedzicki (1988) showed similar sensitivity to antibiotics as *C. Pyogenes* to antibiotics. Most of the examined strains were resistant to erythromycin, ampicillin, streptomycin and nalidixic acid (Myers et al., 1985).

## REFERENCES

- Abd-Alla, A. H. (1990): Studies on some bacteria causing respiratory manifestation in cattle with zoonotic importance. M. V. Sc. Thesis (Microbiology), Faculty of Veterinary Medicine, Zagazig University, Egypt.
- Abdel-Gani, M. El-Seedy, F. R., Shokry, S. and Riad, E. M. (1990): Incidence and bacterial causes of buffalo-calves mortality with respiratory disorders. *Vet. Med. J.*, 38 (2), 233-243.
- Collins, J., K., Jensen, R., Smith, G. H., Flack, D. E., Kersch, R., Bennett, B. W., Jones r. L. and Alexander, A. F., (1988): Association of bovine respiratory syncytial virus with atypical interstitial pneumonia in feedlot cattle. *Amer. J., of vet. Res.* 48 (7), 1045-1049.
- El-Enbaawy, M. (1986): Investigation on some microbiological causes of respiratory affection in buffalo-calves. M. V. Sc. A thesis (Microbiology). Faculty of Vet. Med. Cairo Univ.
- El-Yas, A. H. (1982). Mycological and bacteriological studies on the causes of pneumonia affecting fattening calves. PhD. A thesis, (Medicine). Faculty of Veterinary Medicine Assiut University.
- Finegold, S. M. and Martin, W J (1982): *Diagnostic Microbiology*, 6th edition. C. V. Mosby Co. St. Louis. Toronto, London.
- Fischer, W. (1975): Diagnosis and treatment of possibilities of laryngeal affections in calves. *Dach. Tierarz. Wschr.*, 85, (5), 168-170.
- Galoto, G. B., Mevto, E. and Maserati, R. (1988): Bacterial adherence and respiratory tract disease; a correlation between *Strep. Pyogenes* attachment and recurrent throat



- infection. *Acta Otolaryngol. Suppl.* (Sweden), 454 167-177.
- Garola, Sanda, I., Istrate, N. and Faure, C. (1982): *Haemophilus-like bacteria isolated from calves and lambs. Revta Crest. Animal*, 32, 50-55.
- Hamdy, A. H. and Trapp, A. L. (1967): Investigation of nasal micro flora of feedlot cattle before and after weaning. *Amer. J. of Vet. Res.* 28, 1019-1025.
- Hoerlin, A. B. (1961): Studies on shipping fever of cattle and prevalence of *Pasteurella* spp. in nasal secretion from normal calves. *Ame. J. of Vet. Res.* 22, 470-475.
- Koneman, E. W., Allen, S. D., Dowell, V. R. and Sommers, H. M. (1993): *Colour Atlas and Textbook of Diagnostic Microbiology* 2nd edition, J. B. Lippincott C. New York, London.
- Krieg, N. R. and Holt, J. G. (1984): *Bergey's Manual of Systematic Bacteriology*, Vol 1, William and Wilkins, Baltimore, London.
- Lundberg, C., Heimdahl, A. and Nord, C. E. (1984): Clindamycin in the upper respiratory tract infection. *Scandinavian J. of infect. Dis.* 43, 50-55.
- Magwood, S. E., Barnum, D. A. and Thomas, R. G. (1969): Nasal bacterial flora of calves in healthy and in pneumonic lungs. *Canad J. of Comp Med.* 33, 237-243.
- Myers, L. Shoop, D., Firehammer, B. and Border, (1985): Diseases in calves. *J. Infect, Dis.* 152 1344-1347.
- Nicolas, A., Galaud, C. and Noel, F. (1984): Neonatal diarrhoea. *Med. Vet.*, 160, 107-110.
- Niemela, S. I. Vaatanen, P. and Mentu, Z. (1985): Micro incidence in upper respiratory tract of workers in an industry. *Appl. Environ. Microbiology J.* 50 (1), 10-168.
- Palotay, J. and Newhall, J. (1985): Pneumonia in weaned calves. *J. Amer. Vet. Med. Asso.* 133, 353-39.
- Prasad, B. M. Brivastavo, C. Naragan, K. G. and Prasad, K. (1967): *Pseudomonas pneumonia in calves. Acta V Hung.*, 17, 363 - 369.
- Saleh, R., Morchidi, A. El-Naenaeey, E. Y. and El-Kelbi H. (1993): The sanitary status of El-Qasr slaughterhouse. *Zgazig Vet. J.*, 21, 390-395.
- Singh, S. P., and Singh, N. P. (1980): Milk aspirated pneumonia in calves. *Ind. J. of Animal Sci.* 50, (1) 830-833.
- Wernicki, A., and Rzedzicki, J. (1988): Drugs resistance *E. Coli* isolated from normal calves and those with diarrhoea. *Med. Vet.*, 44, (2), 85-88.
- Wilson, G. S. and Miles, A. A. (1975): *Topley and Wilson Principles of Bacteriology, Virology and immunology* Vol. 1, 6th edition, Edwards Arnold, London.